INCREASED ³H-THYMIDINE INCORPORATION INTO DNA OF ORGAN-CULTURED ADRENAL EXPLANTS FROM RATS INJECTED WITH CORTICOTROPIN AND/OR CYSTEAMINE

E. Sewerynek, M. Szkudliński, A. Lewiński and J. Kunert-Radek

Laboratory of Thyrology, Department of Experimental Endocrinology and Hormone Diagnostic, Institute of Endocrinology, Medical Academy of Łódź, 91-425 Łódź, Poland

Received October 4, 1988

The effect of a single injection of cysteamine /CySH/
- a sulfhydryl substance, known to deplete tissue content of somatostatin /SS/ - on ³H-thymidine incorporation into DNA of rat adrenal explants incubated in vitro was investigated.

It was shown that: 1/ Single in vivo injection of ACTH or of CySH increased ³H-thymidine incorporation into DNA of the organ-cultured adrenals, 2/ Dexamethasone reduced the ³H-thymidine uptake, but that decrease did not attain statistical significance versus controls. © 1988 Academic Press, Inc.

Presence of somatostatin /SS/ receptors has recently been demonstrated in the adrenal cortex of the rat and/or other mammalian species /10/, as well as in human adrenal tumors /i.e., pheochromocytomas and aldosteronomas/ /16/. Some lines of evidence are available, suggesting that SS may be a modulator of the rat adrenal secretory activity /1,2/. This neuropeptide was shown to inhibit the angiotensin II-induced stimulation of aldosterone secretion and the growth of rat adrenal zona glomerulosa cells /11,15/. In turn, chronic SS administration was found to partially reverse the ACTH-enhanced growth of the rat adrenal zona glomerulosa /18/.

Cysteamine /2-mercaptoethylamine, CySH/ is known to deplete the immunoreactive SS content in different organs of the rat and the mouse /23,24/. Beside SS, CySH is also believed to deplete prolactin /Prl/ /12,14/ and growth hormone /GH/ /13/ tissue contents.

In the present study we have attempted to examine the effect of single i.p. injections of ACTH, dexamethasone and/or

CySH on $^3\text{H--thymidine}$ incorporation into DNA of rat adrenals collected 28 h after injection and placed into organ culture.

MATERIALS AND METHODS

Male Wistar rats, weighing 150 ± 20 g each, were donors of adrenals. Twenty eight hours prior to the beginning of incubation the animals were administered one single i.p. injection, as follows:

Group I - controls, 0.9% NaCl, n=8

Group II - ACTH /Synacthen Depot, 1-24 ACTH, CIBA/, 120 U/kg BW, n=8

Group III - Dexamethasone /Decadron phosphate, Merck Sharp and Dohme, The Netherlands/, 2.4 mg/kg BW, n=6

Group IV - Cysteamine /2-mercaptoethylamine, Sigma/, 300 mg/kg BW, n=7
The rats were killed by decapitation. The adrenals, collected

The rats were killed by decapitation. The adrenals, collected from all the animals under sterile conditions, were divided into two equal parts /equatorial cut/ and immediatelly placed in a culture vessel on the surface of a stainless grid for the fluid under the grid to moisten them. The halves of adrenals were incubated for 2 hours in RPMI 1640 medium /Gibco/, containing 2 µCi of 'H-thymidine /Chemapol Prague, Czechoslovakia/ with an addition of 15% fetal calf serum, 20 mM Hepes buffer, penicillin /200 U/ml/ and streptomycin /10 µg/ml/. DNA was extracted as described by Schmidt-Thannhauser /20/, and determined by diphenylamine method /4/. The results were expressed as the mean counts per minute /cpm/ per 1 µg of DNA. Data were statistically analyzed using a one-way analysis of variance /ANOVA/. The significance of differences among the individual groups was estimated with use of Newman-Keuls' test /7/.

RESULTS

The data are illustrated graphically in Figure 1. Corticotropin injection significantly increased $^3\text{H--thymidine}$ incorporation into DNA of adrenal explants /ACTH=305.45 \pm 27.27; controls=73.77 \pm 6.74; $\bar{x}\pm$ SEM/.

Cysteamine administration also induced a significant rise in the tritiated thymidine uptake by adrenal DNA /CySH=147.57 \pm ±15.53/, when compared to the control group.

In contrast, dexamethasone decreased $^3\text{H--thymidine}$ incorporation into DNA of adrenal explants $/43.55\pm8.07/$, but that fall did not attain statistical significance.

DISCUSSION

In the present study, a single in vivo injection of CySH increased H-thymidine incorporation into DNA of the rat adrenal explants

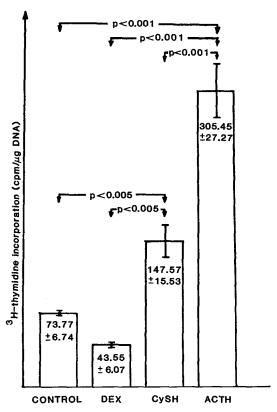


Fig. 1. ³H-thymidine incorporation into adrenal explants in individual incubations; DEX - dexamethasone, CySH - cysteamine.

Data are means ± SEM; p - level of significance.

in organ culture. It is possible that this effect of CySH ensues through a selective depletion of SS tissue content in the adrenals. However, the exact mechanism of CySH action on SS depletion is still unclear. It has been suggested that CySH might act by accelerating the intracellular degradation of immunoreactive SS /24/ or, more likely, through a chemical modification of the disulfide bond which would render the molecule unreactive /17/. Gonzalez-Guijarro et al. /5/ have demonstrated that administration of CySH to rabbits depletes both duodenal mucosa and plasma SS and leads to sensitization or up-regulation of SS binding sites in the duodenum.

Beside SS, CySH is also thought to deplete Prl and GH tissue contents /12,13,14/. Since both GH /26/ and Prl /8/ have been shown to stimulate the adrenal cortex mitogenesis, it is unlikely that tissue depletion of these hormones would stimulate adrenal 3 H-thymidine uptake. On the other hand, an assumption can be

offered that the stimulatory effect of CySH, as obtained in the present study, may be related to the depletion of hypothalamic SS and to the increased pituitary GH release which, as mentioned above, stimulates the adrenal mitosis incidence /26/.

As expected, ACTH injection to rats in vivo significantly increased ³H-thymidine incorporation into DNA of their organ cultured adrenals. It is noteworthy that ACTH in vivo is a potent stimulator of adrenocortical growth /3/ and increases DNA content in adrenals /9/. It was demonstrated by us that ACTH increased the mitotic activity of adrenocortical cells in rats /22,25/ and in mice /21/ in vivo.

In contrast to the stimulating effect of ACTH, we found that dexamethasone - a synthetic glucocorticoid - decreased DNA synthesis in vivo. Our results are in compliance with previous studies reporting the inhibitory effect of dexamethasone on the cortical cell division and on ³H-thymidine incorporation into DNA of the rat adrenal gland /19,27/. The mechanism by which glucocorticoids act on DNA synthesis and cell division remains unclear. It is possible that dexamethasone depresses adrenocortical cell divisions, acting via an inhibition of phospholipase /6/.

In conclusion, CySH-induced increase of adrenal 3H-thymidine uptake, suggests that SS, directly and/or indirectly, may participate in the control of adrenocortical growth. On the other hand, ACTH-induced enhancement and dexamethasone-induced decrease of ³H-thymidine incorporation into the adrenal DNA confirm previous results of other authors.

ACKNOWLEDGMENTS

This work was supported by RMZ-X-23 grant, Contract No. II/08 /A.L. The authors wish to express a deep gratitude to Ms.Ms. Jolanta Kułak, M.Sc., and Jolanta Fryczak, M.Sc., for their excellent technical assistance.

REFERENCES

- 1. Aguilera, G., Harwood, JP., Catt, KJ. /1981/, Nature 292, 262-263.
- 2. Aguilera, G., Parker, DS., Catt, KJ. /1982/, Endocrinology 111. 1376-1384.

- Dallman, MF, /1984-85/, Endocrine Res., 10, 213-242.
 Giles, KW., Myers, A. /1965/, Nature 206, 93-99.
 Gonzalez-Guijarro, L., Lopez-Ruiz, MP., Bodegas, G., Prieto, JC., Arilla, E. /1987/, Exp. Mol. Pathol. 46, 153-158.
 Gryglewski, RJ. /1976/, Pharmacol. Res. Commun. 8, 337-348.

- 7. Hinkle, DE., Wiersma, W., Jurs, SG. /1979/, Applied Statistics for the Behavioral Science. Rand McNally, Chicago.
- 8. Lewiński, A., Sewerynek, E., Webb, S., Esqufino, A., Bartke, A /1988/, Res. Exp. Med. 188, 87-94.
 9. Masui, H., Garren, JD. /1970/, J. Biol. Chem., 245, 2627-2632.
 10. Maurer, R., Reubi, JC. /1986/, Mol. Cell Endocrinol. 45,
- 81-90.
- 11. Mazzocchi, G., Robba, C., Rebuffat, P., Gottardo, Nussdorfer, GG. /1985/, J. Steroid Biochem. 23, 353-356.
- 12. Millard, WJ., Sagar, SM., Martin, JB. /1985/, Fed. Proc. 44, 2546-2550.
- 13. Millard, WJ., Sagar, SM., Badger, TM., Martin, JB. /1983/, Endocrinology 112, 509-517.

 14. Parsons, JA., Peterson, EK., Hartfel, MA. /1984/, Endocrinology 114, 1812-1817.

 15. Rebuffat, P., Robba, C., Mazzocchi, G., Nussdorfer, GG.
- /1984/, J. Steroid Biochem. 21, 387-390.
- 16. Reubi, JC., Maurer, R., von Werder, K., Torchorst, J., Klijn, JGM., Lamberts, SWJ. /1987/, Cancer Res. 47, 551-558.
 17. Rivier, J., Brazeau, P., Vale, W., Guillemin, R. /1975/, J. Med. Chem. 18, 123-126.
- Robba, C., Mazzocchi, G., Nussdorfer, GG. /1986/, Exp. Pathol. 29, 77-82.
 Saez, JM., Morera, AM., Gallet, D. /1977/, Endocrinology 100,
- 1268-1275.
- 20. Schmidt, G., Thannhauser, SJ. /1945/, J. Biol. Chem. 161, 83-89.
- 21. Sewerynek, E., Lewiński, A. J. Pineal Res., in press.
- 22. Sewerynek, E., Lewiński, A., Szkudliński, M., Żerek-Mełeń, G. /1988/, Endokrynol. Pol. in press.
- 23. Sorenson, RL. Grouse, LH., Elde, RP. /1983/, Diabetes 32, 377-379.
- 24. Szabo, S., Reichlin, S. /1981/, Endocrinology 109, 2255-2257. 25. Szkudliński, M. Sewerynek, E., Lewiński, A., Żerek-Mełeń, G., Kułak, J. /1987/, Med, Sci. Res. 15, 657-658.
- 26. Tepperman, I., Engel, FL., Long, CNH. /1943/, Endocrinology 32, 372-4Ó2.
- 27. Wright, NA., Appleton, DR, Morley, AR. /1974/, J. Endocrinol. 62, 527-536.